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Immunomodulation of antigen-specific CD8⁺ T cells by everolimus and mesenchymal stem cells

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Everolimus, a derivative of sirolimus, is a novel inhibitor of mammalian target of rapamycin (mTOR) that is recently used off-label for patients after hematopoietic stem cell transplantation (HSCT) and is useful in reducing cyclosporine-related nephrotoxicity. We demonstrate here that everolimus strongly hampers Cytomegalovirus (CMV)-specific CD8⁺ T cells. Proliferation of CD8⁺ T cells after stimulation with pp65 peptides and the frequency of CMV-specific T cells stained with pp65 tetramers were suppressed by low doses of everolimus in a dose-dependent manner. When we added everolimus at a concentration corresponding to the therapeutic range of the drug (3-8 ng/ml), CD8⁺ T cells were strongly suppressed (by 90%). Beyond proliferation, everolimus had a strong impact on the cell number of CD8⁺ T cells expressing the activation marker CD137 and the secretion of cytokines. In addition we studied the influence of everolimus on T cells after stimulation with staphylococcal enterotoxin B (SEB) or phytohemagglutinin (PHA). Again, proliferation of T cells was inhibited in a dose-dependent manner. As reactivation of CMV plays a pivotal role for the outcome of patients after HSCT, attention must be paid to early detection and pre-emptive treatment of CMV reactivity in patients treated with everolimus.

Mesenchymal stem cells (MSCs) have a potential to differentiate into different cell types such as adipocytes, chondrocytes and muscle cells. Moreover MSCs constitute key players of the immune system. Therefore MSCs have been considered for cellular therapies for patients with graft versus host disease (GVHD) after HSCT. We demonstrated that platelet lysate (PL) -cultured MSCs have similar immunomodulatory capacities when compared to MSCs cultured containing fetal calf serum and cytokines (6S). Both MSCPL and MSC6S showed an immunosuppressive effect on the proliferation of peripheral blood mononuclear cells stimulated with mitogen and mixed lymphocyte reaction (MLR). The immunomodulatory effects of MSCs on CMV-specific CD8⁺ T cells were studied in mixed lymphocyte peptide culture. When co-cultured with either MSCPL or MSC6S, the proliferation of CMV-specific CD8⁺ T cells was inhibited, as well as the frequencies of CD8⁺CMVtetramer⁺ T cells and CD137⁺CMVtetramer⁺ T cells were reduced. Moreover in enzyme-linked immunospot (ELISPOT) assays, MSCs suppressed the number of CMV-specific IFN- γ - and granzyme B-producing cells. In this work, the immunosuppressive potential in terms of inhibition of proliferation, immunophenotype and function of T cells was paradigmatically demonstrated for antigen-specific CD8⁺ T cells.